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Online supplementary material

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Details on PCR conditions

In the Trondheim lab, a separate Polymerase Chain Reaction (PCR) was carried out for each locus with buffer and primer concentrations as described in Jensen et al. (2003). Two-step temperature "touch-down" PCR profiles were used for all loci. Accordingly, we used five cycles at the higher temperature (62° C for Aar8, POCC2; 65° C for Aar5, Ase58, FhU2, Pdou1, Pca3, and Ppi2; 67° C for Aar4 and Ase34), then decreased the annealing temperature by 1° C per cycle (for 10 cycles), followed by 25 cycles at the lower temperature (50° C for Aar8, POCC2; 53° C for Aar5, Ase58, Fhu2, Pdou1, Pca3 and Ppi2; 55° C for Aar4 and Ase34). In the Studenec lab, primers for different loci were combined in a single reaction (i.e. multiplex PCRs) whenever possible. Accordingly, locus Aar4 was amplified in duplex with Ase34, Aar8 with POCC2, and Ase58, FhU2 and Pca3 in a triplex reaction, with the remaining three loci amplified separately. The PCR conditions in the Studenec lab are detailed in Procházka et al. (2008). In the Trondheim lab, each reverse primer was labelled with either 6-FAM (Invitrogen; Aar5, Aar8, Ase34, Pca3, POCC2, Ppi2), VIC (Applied Biosystems; Ase58, FhU2) or NED (Applied Biosystems; Aar4, Pdou1). In the Studenec lab, each forward primer was labelled at the 5' end with one of the following fluorescent dyes (Applied Biosystems): VIC (Aar5, Pca3, Ppi2), NED (Aar8, FhU2, Pdoµ1), PET (Ase34, POCC2) or 6-FAM (Aar4, Ase58). PCR products were pooled in two mixes prior to electrophoresis in

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the Trondheim lab; one mix contained Aar4, Aar5, Ase34, FhU2 and Pca3, the other mix contained Aar8, Ase58, Pdoµ1, POCC2 and Ppi2. In the Studenec lab, PCR products were also pooled in two mixes prior to electrophoresis: one mix included the products of two multiplex reactions (Aar4 and Ase34; Ase58, FhU2 and Pca3), the other contained the products of one duplex (Aar8 with POCC2) and three single reactions (Aar5, Pdoµ1 and Ppi2).

Details on Bayesian clustering programmes

BAPs and STRUCTURE consider multilocus genotypes and attempt to minimize linkage disequilibrium and Hardy-Weinberg disequilibrium by introducing the population structure. In STRUCTURE, we ran five iterations for each K = 1-10 (100 000 burn-in, 500 000 repetitions) using the admixture model, correlated allele frequencies and no prior population information. In BAPs, we performed ten independent runs of spatial clustering of groups of individuals (collected at the same locality). In this program the most likely values of K (in this case the number of clusters of groups) are estimated automatically. TESS performs a maximum likelihood assignment test, which uses geographical information and has been shown to perform very well when F_{ST} values are small (Chen et al. 2007). We followed the suggestions in the software documentation and first ran models without admixture (interaction parameter $\psi = 0.6$) to estimate the number of genetic clusters in the data. For each $K_{\text{max}} = 2-10$, we executed 20 runs and used 20 000 iterations as burn-in and 100 000 subsequent sweeps. Afterwards, as suggested by the authors of the programme, we ran models with admixture using 100 runs for K with the highest likelihood. Twenty runs with the highest likelihood were then averaged in CLUMPP 1.1, a cluster matching and permutation programme (Jakobsson and Rosenberg 2007), and displayed the individual membership coefficients in DISTRUCT 1.1 (Rosenberg 2004).

Table S1. Information on 26 microsatellite loci tested for variability in the reed warbler *Acrocephalus scripaceus*. The loci were tested in 6 individuals; one nestling from each of two separate nests in each of three populations (France, Romania and Denmark). PCR reactions were carried out on a GeneAmp PCR system 9700 (Applied Biosystems). Each 10 μL reaction mix included 0.35 units of AmpliTaq DNA polymerase (Applied Biosystems), 8 μL buffer with final concentration of 20 mM (NH₄)₂SO₄, 75 mM Tris-HCl pH 8.8, 0.15 mg mL⁻¹ DNAse free BSA, 10 mM β-mercaptoethanol, 2.5 mM MgCl₂, 0.14 μM dNTPs (Applied Biosystems), 0.6 μM of each primer, and approximately 20 ng of genomic DNA. All loci were tested using a touch-down PCR profile which started with 4 min at 94 °C followed by 12 cycles of 30 s at 94 °C, 90 s initially at a temperature of 62 °C (then decreasing by one degree every cycle), and 60 s at 72 °C. This was followed by 20 cycles of 30 s at 94 °C, 90 s at 50 °C, and 60 s at 72 °C. Finally there was a final elongation step at 72 °C for 5 min. PCR products were separated by electrophoresis on 6% denaturing acrylamide gels and visualized by silver staining (Bassam et al. 1991).

Locus	Source species	Reference	Repeat motif in source species	Approximate expected allele size	Observed mumber of alleles	Observed allele size*	Description of quality
Aar2 Aar4 Aar4 Aar5 Aar6 Ase18 Ase3 Ase4 Ase64 Ase64 Hru2 Hru2 Hru2 Hru2 Pca3 Pca4 Pca5 Pocc2 Pdol	Great reed warbler Acrocephalus arundinaceus Seychelles warbler Acrocephalus sechellensis Bun swallow Hirundo rustica Barn swallow Hirundo rustica Bun tit Parus caeruleus Blue tit Parus caeruleus Blue tit Parus caeruleus Blue tit Parus caeruleus House sparrow Passer domesticus	Hansson et al. 2000 Richardson et al. 2000 Primmer et al. 1995 Primmer et al. 1995 Primmer et al. 1995 Double et al. 1997 Dawson et al. 2000 Dawson et al. 1997 Neumann and Wetton 1996	(CA), (CA), (CA), (CA), (CA), (CA), (CTTCT), (CTTCT), (CTTTTT), (CTTCT), (CTTCT), (CTTCT), (CTTCT), (CTTCT), (CTTCT), (CTT, (CTT, (CT), (CTCT), (C	132 194 118 115 220 270 270 311 400 412 137 137 170 191 191 191 191 191 191 191 191 191 19	228	100-175 70-80 90-120 225-350 175-300 Long Long 125-175 100-150 150-200 50-75 175-225	Not variable Not variable Good Good Much unspecific amplification Good Not variable Not variable Not variable Good Weak amplification Weak amplification Weak amplification Much unspecific amplification Good Not variable Not variable Good Much unspecific amplification Good Good Good Good Good Good
Pdo4 Pdo5 Pdo10 Ppi2	House sparrow <i>Passer domesticus</i> House sparrow <i>Passer domesticus</i> House sparrow <i>Passer domesticus</i> Magpie <i>Pica pica</i>		(AnGn)n(GAGAGAAA) _x (GAAA) _y (CA) _x (CA) _x	230–500 230 125		200–250	Much unspecific amplification Not variable Clearer if optimized? Good

*To size products a 25 bp Gibco BRL DNA ladder was used in every 25th lane on the 100 lane gel. Sizes were not always noted if the locus was non-variable or there was much unspecific amplification. § This locus was originally named PTC2 in Ellegren 1992.

Table S2. Pairwise F_{ST} values for all 31 reed warbler populations across 10 microsatellite loci (below the diagonal). P-values are given above the diagonal. Values significant after adjusting for multiple comparisons using the false discovery rate approach are given in bold. For population abbreviations see Table 1.

TKM	0.0023 0.2213 0.2404 0.0657 0.1535 0.0013 0.00058 0.00058 0.00058 0.0003 0.0000	
JOA	0.0001 0.0001	0.0100
CYP	0.0015 0.0004 0.0004 0.0009 0.00015 0.00015 0.0001	0.000
Ĕ	0.0134 0.01869 0.02039 0.02037 0.01587 0.037 0.0304 0.0084	-0.0040
HRN	0.00003 0.5353 0.0587 0.0447 0.0447 0.0015 0.0015 0.0015 0.0019 0.0019 0.0013 0.0003 0.0002 0.0003 0.0003 0.0003 0.0003 0.0001 0.0004 0.0003 0	0
HRD	0.002 0.0988 0.0553 0.05125 0.05126 0.0204 0.0053 0.0065 0.0065 0.0065 0.0065 0.0065 0.0065 0.0065 0.0065 0.0065 0.0065 0.0063 0	0.004
SKT	0.1242 0.01567 0.01567 0.01565 0.0034 0.0004 0.0007 0.0015 0.0003 0.0000	0.00.0
HUE	0.0007 0.0066 0.0108 0.0017 0.0017 0.0017 0.0019 0.0001 0.	0.0157
HUC	0.0003 0.1967 0.0068 0.0061 0.0061 0.0001 0.	0.0007
HUW	0.0002 0.02334 0.00395 0.00297 0.02217 0.0227 0.0037 0.0037 0.0037 0.0037 0.0037 0.0037 0.0037 0.0037 0.0037 0.0037 0.0037 0.0003 0.0003 0.0015 0.0015 0.0015 0.0015	-0,0000
BGW	0.0001 0.0146 0.0041 0.0041 0.0041 0.00001 0.0001 0.0001 0.0001 0.0001 0.0001 0.0001 0.0001 0.0001 0.00001 0.00001 0.0001 0.0001 0.0001 0.0001 0.0001 0.0001 0.0001 0.0001 0.0001	0.000
ROD	0.0004 0.0007 0.0007 0.0007 0.0009 0.0009 0.0118 0.0009 0.0009 0.0009 0.0009 0.0009 0.0009 0.0009 0.0009 0.0009 0.0009	-0.0007
ROT	0.0007 0.0007 0.0009 0.0016 0.0016 0.0001	0.010.
PTE	0.0001 0.0001 0.0001 0.0001 0.0001 0.0001 0.0001 0.0001 0.0001 0.0002 0.0003 0.00354 0	0.0121
ESB	0.0001 0.	0.0232
ESG	0.00001 0.0001 0.0001 0.0001 0.0001 0.0001 0.0001 0.0001 0.0015 0.0015 0.0015 0.0015 0.0021 0.0021 0.0021 0.0021 0.0021 0.0021 0.0021 0.0021 0.0021 0.0021 0.0021 0.0021	0.013
ESR	0.00033 0.0495 0.0495 0.0513 0.0513 0.0225 0.0225 0.00186 0.00146 0.00146 0.00146 0.00186 0.00186 0.00186 0.00187 0.00186 0.00187 0.00187 0.00188	0.0137
ESA	0.0282 0.089 0.0556 0.00152 0.00152 0.00194 0.0019 0.0019 0.0011 0.0015 0.0015 0.0015 0.0015 0.0015 0.0015 0.0015 0.0017 0.0018 0.0017 0.0018 0.0018 0.0019 0.0019 0.0019 0.0019 0.0019 0.0019 0.0019 0.0019 0.0019 0.0019 0.0019 0.0019 0.0019 0.0019 0.0019	0.000
ESM	0.00013 0.0005 0.0005 0.00019 0.00019 0.00010	0.010.0
ESE	0.0001 0.0003 0.0003 0.0001 0.0001 0.0002 0.0001 0.0001 0.0003 0.0	0.0000
FLR	0.0002 0.0004 0.0001 0.0001 0.0001 0.0001 0.0002 0.0015 0.	0.012.7
CZS	0.0001 0.0018 0.0311 0.0031 0.0035 0.0005 0.0005 0.0015 0.0155 0.0165 0.	0.0002
UKW	0.00004 0.0002 0.0001 0.0001 0.0001 0.0001 0.0014 0.0023 0.0021 0.0028 0.0021 0.0028 0.0028 0.0021 0.0028 0	0.0171
DKA	0.0766 0.5881 0.1001 0.2271 0.05588 0.1144 0.0158 0.0119 0.0015 0.0015 0.0015 0.0016 0	0.002/
FRB	0.00009 0.4654 0.0169 0.00377 0.00377 0.00137 0.00137 0.00137 0.0013 0.0119 0.0013	0.0042
ΓIA	0.0165 0.04865 0.04877 0.0002 0.0002 0.00074 0.0014 0.0013 0.00013 0.0	0.00.0
CZH	0.00873 0.0137 0.0013 0.0013 0.0013 0.0013 0.0013 0.0014	0.002
PLM	0.0056 0.5101 0.3902 0.0039 0.0039 0.0015 0.0015 0.00145 0.00145 0.0023 0.0023 0.0023 0.0023 0.0023 0.0023 0.0023 0.0023 0.0023 0.0023 0.0023 0.0023 0.0023 0.0023 0.0023 0.0023 0.0023 0.0023	0.0000
DEB	0.00009 0.5044 0.0005 0.0005 0.0005 0.0002 0.0013 0	0.000.0
DEN	0.034 0.0003 0.0003 0.0003 0.0003 0.0003 0.0019 0.0019 0.0019 0.0021 0.0022 0.0023 0.0023 0.0023 0.0023 0.0023 0.0023 0.0033 0.0033	0.00.0
NOR	0.0041 0.0034 0.0089 0.0089 0.0087 0.0017 0.017 0.017 0.0014 0.0021 0.0016 0.00	0.0000
	NOR DEB PLM PLM CZH CZH CZA CZS CZS CZS CZS CZS CZS CZS CZS CZS CZS	INA

P values above the diagonal, **bold significant after adjusting to multiple comparisons** Pairwise Fst values below the diagonal

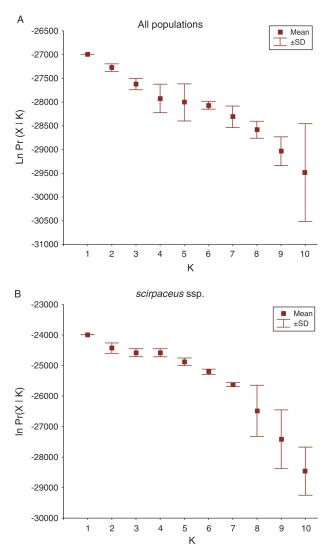


Figure S1. Estimated likelihood for 1–10 genetic population clusters (*K*) in STRUCTURE (100 000 burn-in, 500 000 repetitions) for (A) all populations and (B) *scirpaceus* populations.

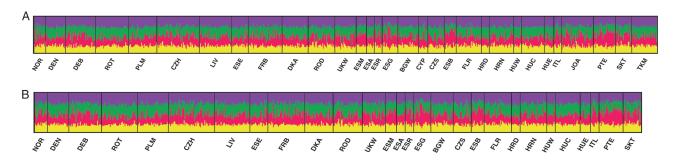


Figure S2. DISTRUCT plot of mean membership coefficients of 20 runs (of 100) with the highest likelihood to four clusters estimated in TESS (admixture, ψ = 0.6) and averaged in CLUMPP for 791 reed warbler individuals from 31 breeding populations (a) and 707 individuals from 28 *scirpaceus* populations (b), respectively. Columns represent individuals, colours assignment probabilities of birds to individual clusters, vertical black bars separate sampling sites.

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